See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/354198763

Short Communication: Silba adipata (Diptera: Lonchaeidae) parasitoids on cayenne pepper (Capsicum frutescens) in Bali, Indonesia

Article *in* Biodiversitas Journal of Biological Diversity · September 2021 DOI:10.13057/biodiv/d220939

CITATION READS 46 1 7 authors, including: Ketut Ayu Yuliadhi I Wayan Supartha Udayana University 19 PUBLICATIONS 14 CITATIONS 83 PUBLICATIONS 98 CITATIONS SEE PROFILE SEE PROFILE Ni Nengah Darmiati I Kadek Wisma Yudha Udayana University 1 PUBLICATION 1 CITATION 15 PUBLICATIONS 17 CITATIONS SEE PROFILE SEE PROFILE

Some of the authors of this publication are also working on these related projects:

Liriomyza View project

New Record of Silba adipata McAlpine (Diptera: Lonchaeidae) as a Pest of White Chili Fruit in Bali of Indonesia View project

**BIODIVERSITAS** Volume 22, Number 9, September 2021 Pages: 3929-3935

# Short Communication: Silba adipata (Diptera: Lonchaeidae) parasitoids on cayenne pepper (Capsicum frutescens) in Bali, Indonesia

# KETUT AYU YULIADHI<sup>1</sup>, I WAYAN SUPARTHA<sup>1,•</sup>, NI NENGAH DARMIATI<sup>1</sup>, ALPREDO BANGUN<sup>2</sup>, I KADEK WISMA YUDHA<sup>3</sup>, I WAYAN EKA KARYA UTAMA<sup>3</sup>, PUTU ANGGA WIRADANA<sup>4</sup>

<sup>1</sup>Laboratory of Integrated Pest Management, Faculty of Agriculture, Universitas Udayana. Jl. P.B. Sudirman, Denpasar 80234, Bali, Indonesia
<sup>2</sup>Program of Agriculture Graduate Program, Faculty of Agriculture, Universitas Udayana. Jl. P.B. Sudirman, Denpasar 80234, Bali, Indonesia
<sup>3</sup>Dry Land Agriculture Graduate Program, Faculty of Agriculture, Universitas Udayana. Jl. P.B. Sudirman, Denpasar 80234, Bali, Indonesia
<sup>4</sup>Program of Biology, Faculty of Health, Science and Technology, Universitas Dhyana Pura. Jl. Raya Padang Luwih, North Kuta, Badung 80361, Bali, Indonesia. Tel.: +62-822-4796-6490, Fax.: +62-361-701907

Manuscript received: 28 June 2021. Revision accepted: 26 August 2021.

**Abstract.** *Yuliadhi KA, Supartha IW, Darmiati NN, Bangun A, Yudha IKW, Utama IWEK, Wiradana PA. 2021.* Silba adipata (*Diptera: Lonchaeidae*) parasitoids on cayenne pepper (Capsicum frutescens) in Bali, Indonesia. Biodiversitas 22: 3929-3935. The black fig fly (*Silba adipata*) is newly emerging pests that attack cayenne pepper (*Capsicum frutescens* L.) plants in Bali Province, Indonesia. This study aimed to determine the parasitoid type, community structure, distribution, and parasitization rate of the parasitoids of *S. adipata* on cayenne pepper in Bali. The sampling method employed in this study involves purposive sampling techniques on 100-150 pieces of cayenne pepper infested with *S. adipata* at each location point, with the emerging parasitoids identified morphologically. Three types of parasitoid species were associated with *S. adipata*, which include *Asobara japonica* Belokobylskij, *Fopius arisanus* (Sonan), and *Diachasmimorpha longicaudata* (Ashmead). The parasitoid community structure associated with *S. adipata* had a low abundance index, low diversity index, and a moderate dominance index. Based on the distribution of the three parasitoids, *D. longicaudata* is distributed evenly in Bali, whereas *F. arisanus* was not found in Jembrana . Conversely, *A. japonica* was only found in Badung, Bangli, Gianyar, and Klungkung. As summary, *D. laungicaudata* had the highest parasitization rate compared to the *F. arisanus* and *A. japonica*. *D. longicaudata* is a potential parasitoid, which can be studied for the future biological control of *S. adipata*.

Keywords: Biological control, Black Fig Fly, Braconidae, geographical distribution, Silba adipata

## **INTRODUCTION**

Cayenne pepper (*Capsicum frutescens* L.) is a horticultural commodity that is widely cultivated in Indonesia, which corresponds to an increase in national cayenne pepper production increased from 800,484 tons in 2014 to 1,335,608 tons in 2018 (Indriani et al. 2020). However, the production of white cayenne pepper in Bali Province, Indonesia has fluctuated since it increased from 28,440 tons in 2014 to 38,358 tons in 2016 and then decreased to 31,655 tons in 2018 (BPS 2019).

The decreasing cayenne pepper production can be caused by abiotic and biotic factors (Borowski et al. 2019). Also, the cayenne pepper plantation in Bogor (West Java) has been infested by a new pest-black fig fly (BFF) *Silba capsicarum* (Diptera: Lonchaeidae) (MacGowan and Rauf 2019). According to previous study, 911 adults of *S. capsicarum* were recovered from 281 chilli peppers (MacGowan and Rauf 2019). *Silba adipata* McAlpine is a new pest that infests cayenne pepper in Bali, and its morphology differs from that of the *S. capsicarum* species previously reported in Bogor, West Java. More so, one of the morphological differences is a puncture symptom from the ovipositor a brown dot on the immature fruit surface (Merta 2019).

Due to the lack of bioecological information on pest management, farmers use broad-spectrum. However, this tactic provides a negative impact on the environment and the health of consumers and farmers themselves (Lee et al. 2013). Alternative strategy such as parasitoids is promising in suppressing pest infestations peaks (Bezerra et al. 2021). Naturally, interspecific competition between parasitoids may occur if two or more species attack the same host species in order to compete for resources (Harvey et al. 2013), or may have an additive effect. This competition may also allow for the emergence of fundamental potential in biological pest control strategies, such as the displacement of one parasitoid species by another or a reduction in overall effectiveness for the target pest (Wang et al. 2016). This competition is also known as "intra and interspecific competition" (Harvey et al. 2013). As a result, understanding parasitoid types and levels of parasitization is critical in biological control programs against specific pest (Mi et al. 2021). This study investigated the species, community structure, distribution, and parasitization rate of parasitoids against the black fig fly, S. adipata on cayenne pepper in Bali Province, Indonesia.

## MATERIALS AND METHODS

## **Study location**

The study was conducted from February to April 2020 in 8 districts and 1 city in Bali Province, Indonesia as shown in Figure 1. Laboratory research was conducted at the Integrated Pest and Disease Management Laboratory (IPMLab) at the Faculty of Agriculture, Udayana University, Bali.

## Sampling method

The sampling location in each district and city was determined based on the cayenne pepper data provided on the Bali Province BPS website (bali.bps.go.id). 100-150 cayenne pepper samples infested by *S. adipata* were taken purposively from each location point, and then stored in sterile plastic bags, labeled, and brought to the laboratory for observation.

### Maintenance procedure

The white cayenne pepper was maintained by placing it in a transparent plastic container with a 6 cm lower diameter, 9 cm upper diameter, and 12.5 cm height. The container was filled with 20 g of sandy soil, which was used as a medium for pupation. The humidity level was maintained by spraying with water, and infected cayenne pepper was placed in the container and covered with gauze. Each container was covered with gauze, labeled with the field data, and placed in rearing containers until emergence of flies or parasitoids.

#### **Parasitoid identification**

The identification of parasitoid type was carried out by analyzing the morphological characteristics of insects according to the key determination of Sharkey (1992) and Wharton and Lopez-Martinez (2000).

## The community structure

To determine the community structure, measurements on species diversity index, abundance index and dominance index of parasitoids were applied.

The diversity index was measured using an index developed by Shannon and Wiener (H<sup> $\circ$ </sup>) (Magurran 2005), with the following equation:

$$H' = -\sum pi. ln. pi$$

Where:

*H*': Shannon Wiener Index

*ni* : Number of species individuals

N : Total number of individuals

The diversity index was grouped into three categories, namely H'<1.5 (low diversity), H=1.5-3.5 (moderate diversity), and H' > 3.5 (high diversity).

The abundance index was measured using the Margalef index (Magurran 2005).

$$Rl = \frac{S-1}{\ln N}$$

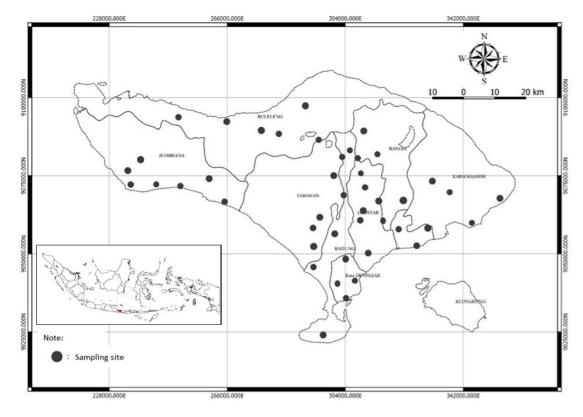


Figure 1. Sampling site in Bali, Indonesia

Where:

R1: Abundance indexS: Number of species foundln: Nature logarithmN: Total number of individualsValues:R1 < 3.5 = 10wR1 > 3.5 - <5.0 = moderateR1 > 5.0 = high

The dominance index was measured using the Menheinick index (Magurran 2005).

$$D = \sum \left( \frac{ni(ni-1)}{N(N-1)} \right)$$

Where:

D : Dominance index

N: Total number of individuals

ni : Number of i-species individuals

Values:  $D = \le 0.00-0.30$ : low D = > 0.30-0.60: moderate  $D = \ge 0.60-1.00$ : high

## **Parasitization rate**

The parasitization rate was determined by counting the number of flies and adult parasitoids seen in each observation. This was determined using the following formula:

$$P = \frac{\sum adult \ parasitoid \ A}{\sum adult \ S. adipata + \sum adult \ parasitoids \ found} \times 100\%$$

Where:

P = parasitization rate (%)  $\sum \text{ adult parasitoid A} = \text{number of parasitoids}$  $\sum \text{ adult S. adipata} = \text{number of adult S. adipata}$ 

#### Data analysis

Morphological characteristics, are presented as images, whereas the community structure and parasitization rate were tabulated and analyzed using Ms. Excel 2019 (Microsoft, USA) and presented in the form of tables and graphs.

#### **RESULTS AND DISCUSSION**

There were three species of parasitoid associated with *S. adipata*, which include *Asobara japonica* Belokobylskij, *Fopius arisanus* Sonan, and *Diachasmimorpha longicaudata* Ashmead.

#### **Descriptions and general information**

Asobara japonica is distinguished by morphological characteristics such as an antenna that is longer than the body size. As shown in Figure 2A, the body length is 3.87

mm and the antenna length is 3.98 mm. These species have similarities with the parasitoid specimen discovered by Suriani (2020), which has a black ovipositor measuring 2.66 mm in length. Figure 2B shows, species 1 with a darkbrown head and a thorax with a reddish-brown propodeum.

Asobara japonica is a larva-pupae endoparasitoid, with a wide host range, mainly found in larvae of the genus Drosophila species, that plays an important role in controlling Drosophila melanogaster and Drosophila suzukii (Matsumura) (Zhang et al. 2020). This parasitoid can kill the host, causing the parasitoid larvae to die as well (Prevost et al. 2012). Interestingly, this parasitoid can secrete a natural toxin with an atypical effect on the host causing permanent paralysis followed by the death of D. melanogaster larvae (Mabiala-Moundoungou et al. 2010). Also, Hymenoptera parasitoids use volatile compounds to locate their hosts, particularly in microhabitat locations. According to the findings of Biondi et al. (2017), the olfactory response of adult Asobara japonica illustrates its innate interest or plasticity in exploiting volatile compounds from various fruits infested by the invasive pest Drosophila suzukii. The findings of these parasitoid larvae are very helpful in improving protocols for the maintenance and evaluation of D. suzukii infesting cherry, blackberry, or strawberry especially in smaller host sizes (Wang et al. 2020; Wang et al. 2021).

*Fopius arisanus* has a black-brown body with a yellowish-brown abdomen. Furthermore, it has a body length of 4.55 mm, and an antenna length of 5.32 mm, with black color. Also, the ovipositor is 3.46 mm long as shown in Figure 3A. The abdomen is yellowish-brown with a second tergum in a stripe pattern as shown in Figure 3B.a. Furthermore, the mesoscutum is equipped with a notauli shaped like a necklace as shown in Figure 3C.a and Figure 3C.b.

*Fopius arisanus* is an egg-larva parasitoid that has been widely developed for its potential in biological control programs against the main tephritid pests in fruit commodities, including *Bactrocera dorsalis* (Hendel) and *Ceratitis capitata* (Wiedemann) (Groth et al. 2016). In addition, it was also able to parasitize one-day-old *Ceratitis cosyara* eggs (Karlsson et al. 2018). Furthermore, *F. arisanus* has a haplodiploid mating system, which involves mating with male offspring from unfertilized eggs and females from eggs fertilized by male sperm. On a larger scale, the release of *F. arisanus* was carried out due to its polyphagous nature and bulk compatibility in controlling fruit fly infestations. (Vargas et al. 2012a).

*Diachasmimorpha longicaudata*, which exhibit a body length of 4.69 mm, is the subject of Specimen 3. It has a bright orange body with a black-brown tarsus. Also, as shown in Figure 4A.a, the antenna is 6.4 mm long and has a dark ovipositor that is 5.5 mm or longer than the body size. As illustrated in Figure 4B, the mesoscutum lacks notauli, whereas the propodeum is orange.

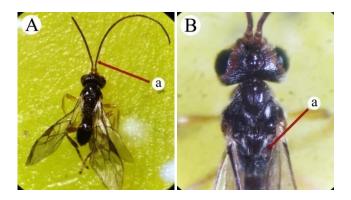
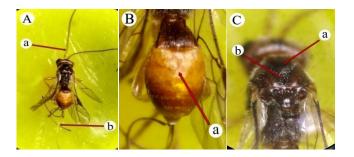
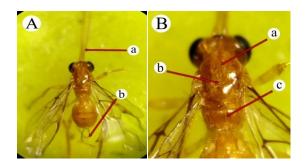


Figure 2. Specimen 1 of *A. japonica*. A. overall appearance (a. antennae) and B. thorax (a. propodeum)



**Figure 3.** *Fopius arisanus.* A. overall appearance (a. antennae; b. ovipositor). B. Abdomen (a. tergum II). C. Thorax (a. Mesoscutum; b. Notauli).



**Figure 3.** Specimen 3, *Diachasmimorpha laungicaudata*. A. overall appearance (a. antennae; b. ovipositor). B. Thorax (a. Mesoscutum; b. no notauli; c. propodeum)

## Silba adipata versus parasitoids in Bali

All recovered Bracondidae species had different diversity, abundance, and dominance index values. The parasitoids associated with *S. adipata* had a "low" abundance index (H' < 3.5) in all districts and city in Bali with an average abundance index ranging between 0.00-0.76. The parasitoids had a "low" diversity index less than (H' < 1.0) in all districts and city in Bali with an average diversity index of 0.00-1.02. Meanwhile, there were various dominance index values. The "moderate" dominance index (0.30 < D < 0.60) was found in Badung, Bangli, Gianyar, Karangasem, Klungkung and Denpasar

*Diachasmimorpha longicaudata* (Hymenoptera: Braconidae) is a solitary coinobiont endoparasitoid wasp that lays its eggs in fruit fly larvae where it grows to adulthood (Koskinioti et al. 2020). This is considered as one of the potentials for good biocontrol as an augmentative application against Tephritidae fruit flies. For example, *D. longicaudata* controlling populations of *C. capitata* (Sánchez et al. 2016; Harbi et al. 2018), *B. dorsalis* (Vargas et al. 2012b), *Bactrocera oleae* (Rossi) (Sime et al. 2008), and *B. zonata* (Saunders) (Andleeb et al. 2010). Similarly, *D. longicaudata* is an effective biological control agent against medflies, which infests peach, as well as sour and sweet orange (Suárez et al. 2019).

with an average value of 0.37-0.55. While the high dominance index value (D>0.60) was found in Buleleng, Jembrana, and Tabanan (0.63-1.00) (Table 1).

Braconidae parasitoids are parasitoids that attack larvae in fruit and are a viable potential for biological control for several reasons. First, natural enemies were able to suppress the population of *S. adipata* cayenne pepper in the field. Second, ecosystems that are rarely touched by human activities and production inputs in agricultural cultivation are actually considered to have high levels of biodiversity in some areas (Wahyuni et al. 2017). Third, genetic resistance, sex ratio, and parasitoid adaptability are also able to determine this (Supartha et al. 2021).

#### Distribution of S. adipata parasitoids in Bali

According to the results, *D. laungicaudata* and *F. arisanus* are both associated with *S. adipata* and were found in all districts and city throughout Bali. *F. arisanus* was not found in Jembrana, whereas *A. japonica* was only found in Badung, Bangli, Gianyar, and Klungkung as shown in Figure 4. In addition, several factors influence parasitoid distribution, including biotic and abiotic factors (Supartha et al. 2020; Yudha et al. 2020).

## The parasitization rate of S. adipata parasitoids in Bali

The following parasitization rate by D. longicaudata per locality was obtained: Karangasem, Tabanan, Jembrana, and Denpasar as 28.18%, 26.56%, 23.21%, and 11.53%, respectively. While that of Klungkung, Buleleng, Bangli, Gianyar and Badung were 10.10%, 6.06%, 5.54%, 4.82% and 4.17%, respectively. Furthermore, the parasitization rate of F. arisanus in Karangasem, Gianyar, Denpasar, Bangli, and Buleleng was 16.06%, 7.71%, 5.71%, 3.81%, and 3.57%, respectively. While that of Klungkung, Tabanan, Badung, and Jembrana were 3.03%, 2.46%, 2.04%, and 0.00%, respectively. The parasitization rate by A. japonica in Bangli, Klungkung, Badung, and Gianyar was 1.37%, 1.01%, 0.86%, and 0.71%, respectively, whereas, Buleleng, Jembrana, Karangasem, Tabanan, and Denpasar were all at 0.00%, as shown in Figure 6.

Table 1. Community s	tructure of parasitoid	Silba adipata in Bal	i, Indonesia

Spesies	District/ City								
	Badung	Bangli	Buleleng	Denpasar	Gianyar	Jembrana	Karangasem	Klungkung	Tabanan
D. longicaudata	60	43	17	4	25	4	15	10	18
F. arisanus	31	34	1	2	80	0	7	3	6
A. japonica	13	15	0	0	17	0	0	1	0
N parasitoid	104	92	18	6	122	4	22	14	24
N S. adipata	1314	878	322	29	611	14	26	90	509
S parasitoid	3	3	2	2	3	1	2	3	2
S S. adipata	1	1	1	1	1	1	1	1	1
R1	0,43	0,44	0,35	0,56	0,42	0,00	0,32	0,76	0,31
H'	0,94	1,02	0,21	0,64	0,88	0,00	0,63	0,76	0,56
D	0,43	0,37	0,89	0,47	0,49	1,00	0,55	0,53	0,63

Note: N: Individual abundance, S: Species abundance, R1: Abundance Index, H': Diversity index, D: Dominance Index.

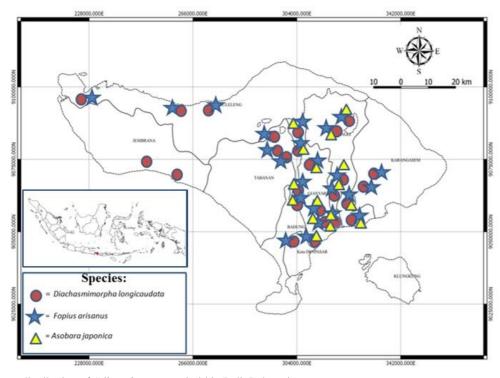


Figure 5. The map distribution of Silba adipata parasitoid in Bali, Indonesia

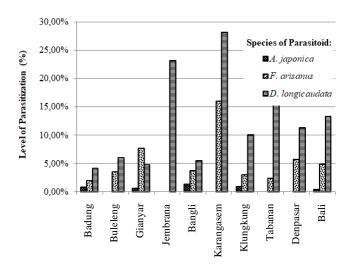


Figure 6. Parasitization rate of three parasitoids (Braconidae) on *Silba adipata* in Bali, Indonesia

In the present study, *D. longicaudata* had the highest potential to control *S. adipata* infestations in cayenne pepper in Bali. The parasitization rate tends to be directly proportional to the host population, and vice versa since the host population decreases with the number of parasitoids (Cusumano and Volkoff. 2021). Meanwhile, preference factors for other types of host prey, as well as demographic changes, can influence the natural decrease in parasitism rates (Papkou et al. 2016). Furthermore, competition between species within the same host promotes the survival of other types of parasitoids (Yang et al. 2013).

Biological control of *S. adipata* using parasitoids is a strategic effort through the introduction of exotic parasitoids (classical biological control) and alternative uses of ecologically based insecticides (Sow et al. 2019). Keseluruhan, parasitoid ini adalah endoparasitoid soliter yang mampu menyerang bagian telur/larva (Wang et al. 2021).

*Diachasmimorpha laungicaudata* had the highest parasitization rate of 28.18%, compared with *F. arisanus* and *A. japonica* (16.06 % and 1.37%, respectively). Overall, our results reveal preliminary information about the parasitoid diversity associated with the new invasive pest *S.ilba adipata* in Cayenne Pepper in Bali. Further studies are still needed to evaluate the efficacy of release of exotic-parasitoids to control *S. adipata*.

## ACKNOWLEDGEMENTS

The authors would like to thank the Head of the Integrated Pest Management Laboratory at Udayana University, Bali, Indonesia, for providing facilities and funding assistance.

## REFERENCES

- Andleeb S, Shahid MS, Mehmood R. 2010. Biology of Parasitoid Aganaspis daci (Weld) (Hymenoptera: Eucoilidae). Pak J Sci Ind Res 53: 201-204
- BPS. 2019. Produksi Tanaman Cabai Rawit Menurut Provinsi, 2014-2018, Badan Pusat Statistik dan Direktorat Jenderal Hortikultura, Jakarta. [Indonesian]
- Bezerra JIM, Molter A, Rafikov M, Frighetto DF.2021. Biological control of the chaotic sugarcane borer-parasitoid agroecosystem. Ecol Model 450: 109564. DOI: 10.1016/j.ecolmodel.2021.109564
- Biondi A, Wang X, Miller JC, Miller B, Shearer PW, Zappalà L, Siscaro G, Walton VW, Hoelmer KA, Daane KM. 2017. Innate olfactory responses of *Asobara japonica* toward fruits infested by the invasive Spotted Wing *Drosophila*. J Insect Behav 30: 495-506.
- Borowski RGV, Zimmer KR, Leonardi BF, Trentin DS, Silva RC, Barros MPD, Macedo AJ, Gnoatto CB, Gosmann G, Zimmer AR. 2019. Red pepper *Capsicum baccatum*: source of antiadhesive and antibiofilm compounds against nosocomial bacteria. Industr Crop Prod 127: 148-157. DOI: 10.1016/j.indcrop.2018.10.011.
- Cusumano A, Volkoff A-N. 2021. Influence of parasitoid-associated viral symbionts on plant-insect interactions and biological control. Curr Opin Insect Sci 44: 64-71. DOI: 10.1016/j.cois.2021.03.009.
- Groth MZ, Loeck AE, Nörnberg SD, Bernardi D, Nava DE. 2016. Biology of *Fopius arisanus* (Hymenoptera: Braconidae) in two species of fruit flies. J Insect Science 16 (1): 96. DOI: 10.1093/jisesa/iew087
- Harbi A, Beitia F, Ferrara F, Chermiti B, Sabater-Muñoz B. 2018. Functional response of *Diachasmimorpha longicaudata* (Ashmead) over *Ceratitis capitata* (Wiedemann): Influence of temperature, fruit location and host density. Crop Protect 109: 115-122. DOI: 10.1016/j.cropro.2018.03.010.
- Harvey JA, Poelman EH, Tanaka T. 2013. Intrinsic inter-and intraspecific competition in parasitoid wasps. Ann Rev Entomol 58 (1): 333-351. DOI: 10.1146/annurev-ento-120811-153622
- Indriani R, Darma R, Musa Y, Tenriawaru AN, Arsyad M. 2020. Policy design of cayenne pepper supply chain development. Bulgarian J Agric Sci 26 (3): 499-506.
- Ives AR, Klug JL, Gross K. 2000. Stability and species richness in complex communities. Ecol Lett 3 (5): 399-411. DOI: 10.1046/j.1461-0248.2000.00144.x.
- Karlsson MF, Souza EO.d, Ayelo PM, Zannou JA, Mègnigbèto GSB, Bokonon-Ganta AH. 2018. Interspecific competition between egg parasitoids: Native *Fopius caudatus* and exotic *Fopius arisanus*, in Ceratitis cosyra. Biol Contr 117: 172-181. DOI: 10.1016/j.biocontrol.2017.11.010.
- Koskinioti P, Ras E, Augustinos AA, Beukeboom LW, Mathiopoulos KD, Caceres C, Bourtzis K.2020. The impact of fruit fly gut bacteria on the rearing of the parasitic wasp *Diachasmimorpha longicaudata*. Entomologia Experimentalis et Applicata 168 (6-7): 541-559. DOI: 10.1111/eea.12936.
- Lashkari-bod A, Zebitz, CPW. 2012. Diversity and abundance of parasitoids in organic apple orchards in Baden-Württemberg. In

Conference: 16th Intl. Conf. on Organic Fruit-Growing At: Stuttgart-Hohenheim, Germany. Stuttgart-Hohenheim, Germany.

- Lee D-H, Short BD, Joseph SV, Bergh JC, Leskey TC. 2013. Review of the Biology, Ecology, and Management of *Halyomorpha halys* (Hemiptera: Pentatomidae) in China, Japan, and the Republic of Korea. Environ Entomol 42 (4): 627-641. DOI: 10.1603/EN13006.
- Mabiala-Moundoungou ADN, Doury G, Eslin P, Cherqui A, Prévost G. 2010. Deadly venom of *Asobara japonica* parasitoid needs ovarian antidote to regulate host physiology. J Insect Physiol 56 (1): 35-41. DOI: 10.1016/j.jinsphys.2009.09.001
- MacGowan I, Rauf A. 2019. Silba capsicarum (Diptera: Lonchaeidae), a newly recognized pest of chilli pepper in Java. J Asia-Pac Entomol 22 (1): 83-86. DOI: 10.1016/j.aspen.2018.12.003
- Magurran AE. 2005. Measuring Biological Diversity. Blackwell Publishing. Oxford, UK.
- Merta IN. 2019. Life History of Black Fly *Silba adipata* (Diptera: Lonchaeidae) as a New Pest of White Caba Rawit Fruit (*Capsicum frutescens* L). [Thesis]. Udayana University, Denpasar. [Indonesian]
- Mi Q, Zhang J, Haye T, Zhang B, Zao C, Lei Y, Li D, Zhang F. 2021. Fitness and interspecific competition of *Trissolcus japonicus* and *Anastatus japonicus*, egg parasitoids of *Halyomorpha halys*. Biol Contr 152: 104461. DOI: 10.1016/j.biocontrol.2020.104461
- Papkou A, Gokhale CS, Traulsen A, Schulenburg H. 2016. Host-parasite coevolution: why changing population size matters. Zoology. 119 (4): 330-338. DOI: 10.1016/j.zool.2016.02.001
- Prevost G, Aslin P, Cherqui A, Moreau S, Doury G. 2012. Chapter 21-When Parasitoids Lack Polydnaviruses, Can Venoms Subdue the Hosts? The Case Study of *Asobara* Species. Parasitoid Viruses. 2012: 255-266. DOI: 10.1016/B978-0-12-384858-1.00021-7.
- Qian H, Jin Y.2016. An updated megaphylogeny of plants, a tool for generating plant phylogenies and an analysis of phylogenetic community structure. J Plant Ecology 9 (2): 233-239. DOI: 10.1093/jpe/rtv047
- Sharkey MJ. 1992. Cladistics and tribal classification of the Agathidinae (Hymenoptera: Braconidae). J Nat Hist 26 (2): 425-447. DOI: 10.1080/00222939200770251
- Sime KR, Daane KM, Wang XG, Johnson MW, Messing RH. 2008. Evaluation of *Fopius arisanus* as a biological control agent for the olive fruit fly in California. Agric For Entomol 10 (4): 423-431. DOI: 10.1111/j.1461-9563.2008.00401.x
- Sow A, Brévault T, Benoit L, Chapuis M-P, Galan M, Coeur d'acier A, Delvare G, Sembène M, Haran J. 2019. Deciphering host-parasitoid interactions and parasitism rates of crop pests using DNA metabarcoding. Nature (Scientific Reports). 9 (1): 3646. DOI: 10.1038/s41598-019-40243-z
- Suárez L, Biancheri MJB, Sánchez G, Murúa F, Funes CF, Kirschbaum DS, Molina D, Laría O, Ovruski SM. 2019. Effects of releasing two Diachasmimorpha longicaudata population lines for the control of *Ceratitis capitata* infesting three key host fruit species. Biological Control. 133: 58-65. DOI: 10.1016/j.biocontrol.2019.03.011
- Supartha IW, Yudha IKW, Wiradana PA, Susila, IW. 2020. Response of parasitoids to invasive pest *Phenacoccus manihoti* Matile-Ferrero (Hemiptera: Pseudococcidae) on cassava crop in Bali, Indonesia. Biodiversitas 21 (10): 4543-4549 DOI: 10.13057/biodiv/d211011
- Supartha IW, Sunari, AAAAS, Krisna IGPB, Yudha IKW, Wiradana PA. 2021. Invasion, Population Development and Attack intensity of the fall armyworm *Spodoptera frugiperda* J.E. Smith (Lepidoptera: Noctuidae) on two variety corn in Serongga Village, Gianyar Regency, Bali, Indonesia. Technology Report of Kansai University. 10 (1): 645-654.
- Suriani N. 2020. Eksplorasi Potensi Spesies Parasitoid Braconidae untuk Rintisan Pengendalian Hayati Hama Baru Silba sp. (Diptera: Lonchaeidae) Pada Cabai Rawit Putih (*Capsicum frutescens* L.) [Tesis]. Udayana University. [Indonesian].
- Vargas RI, Leblanc L, Harris EJ, Manoukis NC.2012. Regional Suppression of Bactrocera Fruit Flies (Diptera: Tephritidae) in the Pacific through Biological Control and Prospects for Future Introductions into Other Areas of the World. Insects. 3 (3): 727-742. DOI: 10.3390/insects3030727.
- Vargas RI, Leblanc L, Putoa R, Piñero JC.2012. Population dynamics of three *Bactrocera* spp. fruit flies (Diptera: Tephritidae) and two introduced natural enemies, *Fopius arisanus* (Sonan) and *Diachasmimorpha longicaudata* (Ashmead) (Hymenoptera: Braconidae), after an invasion by *Bactrocera dorsalis* (Hen. Biol Contr 60 (2): 199-206. DOI: 10.1016/j.biocontrol.2011.10.012.

- Wahyuni S, Supartha IW, Ubaidillah R, Wijaya IN. 2017. Parasitoid community structure of leaf miner Liriomyza spp. (Diptera: Agromyzidae) and the rate of parasitization on vegetable crops in Lesser Sunda Islands, Indonesia. Biodiversitas 18 (2): 593-600. DOI: 10.13057/biodiv/d180221
- Wang X.-G, Kaçar G, Biondi A, Daane KM. 2016. Foraging efficiency and outcomes of interactions of two pupal parasitoids attacking the invasive spotted wing drosophila. Biol Contr 96: 64-71. DOI: 10.1016/j.biocontrol.2016.02.004.
- Wang X, Lee JC, Daane KM, Buffington ML, Hoelmer KA. 2020. Biological control of *Drosophila suzukii*. CAB Rev Perspect Agric Vet Sci Nutr Nat Resour 15 (54): 1-18. DOI: 10.1079/PAVSNNR202015054.
- Wang X, Biondi A, Nance A H, Zappalà L, Hoelmer K A and Daane K M. 2021 Assessment of Asobara japonica as a potential biological control agent for the spotted wing drosophila, Drosophila suzukii. Entomol Gen 41: 1-12. DOI: 10.1127/entomologia/2020/1100.
- Wang X, Ramadan MM, Guerrieri E, Messing RH, Johnson MW, Daane KM, Hoelmer KA. 2021 Early-acting competitive superiority in opine parasitoids of fruit flies (Diptera: Tephritidae): Implications for

biological control of invasive tephritid pests. Biol Contr 162: 104725. DOI: 10.1016/j.biocontrol.2021.104725.

- Wharton R. Lopez-Martinez V. 2000. A new species of *Triaspis haliday* (Hymenoptera: Braconidae) parasitic on the pepper weevil, *Anthonomus eugenii* Cano (Coleoptera: Curculionidae). Proceedings of the Entomological Society of Washington, Washington DC.
- Yang S, Duan JJ, Lelito J, Driesche RV. 2013. Multiparasitism by *Tetrastichus planipennisi* (Hymenoptera: Eulophidae) and *Spathius agrili* (Hymenoptera: Braconidae): Implication for biological control of the emerald ash borer (Coleoptera: Buprestidae). Biol Contr 65 (1): 118-123. DOI: 10.1016/j.biocontrol.2012.09.010.
- Yudha IKW, Supartha IW, Susila IW. 2020. Karakteristik morfologi dan kelimpahan populasi parasitoid (Hymenoptera) pada hama invasif kutu putih ubi kayu, *Phenacoccus manihoti* Matile-Fererro (Hemiptera: Pseudococcidae) di Bali. Agrotrop 10 (2): 178-189. DOI: 10.24843/ajoas. 2020.v10.i02.p07. [Indonesian]
- Zhang X, Li C, Pan Z, Zhu J, Wang Z, Shi M, Chen X, Huang J. 2020. The complete mitochondrial genome of *Asobara japonica* (Hymenoptera: Braconidae). Mitochondrial DNA Part B 5 (2): 1279-1281. DOI: 10.1080/23802359.2020.1732238.